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## FORMULATION DEVELOPMENT OF ENTERICALLY **PROTECTED SPRAY DRIED DISPERSIONS OF ADRULIPASE**

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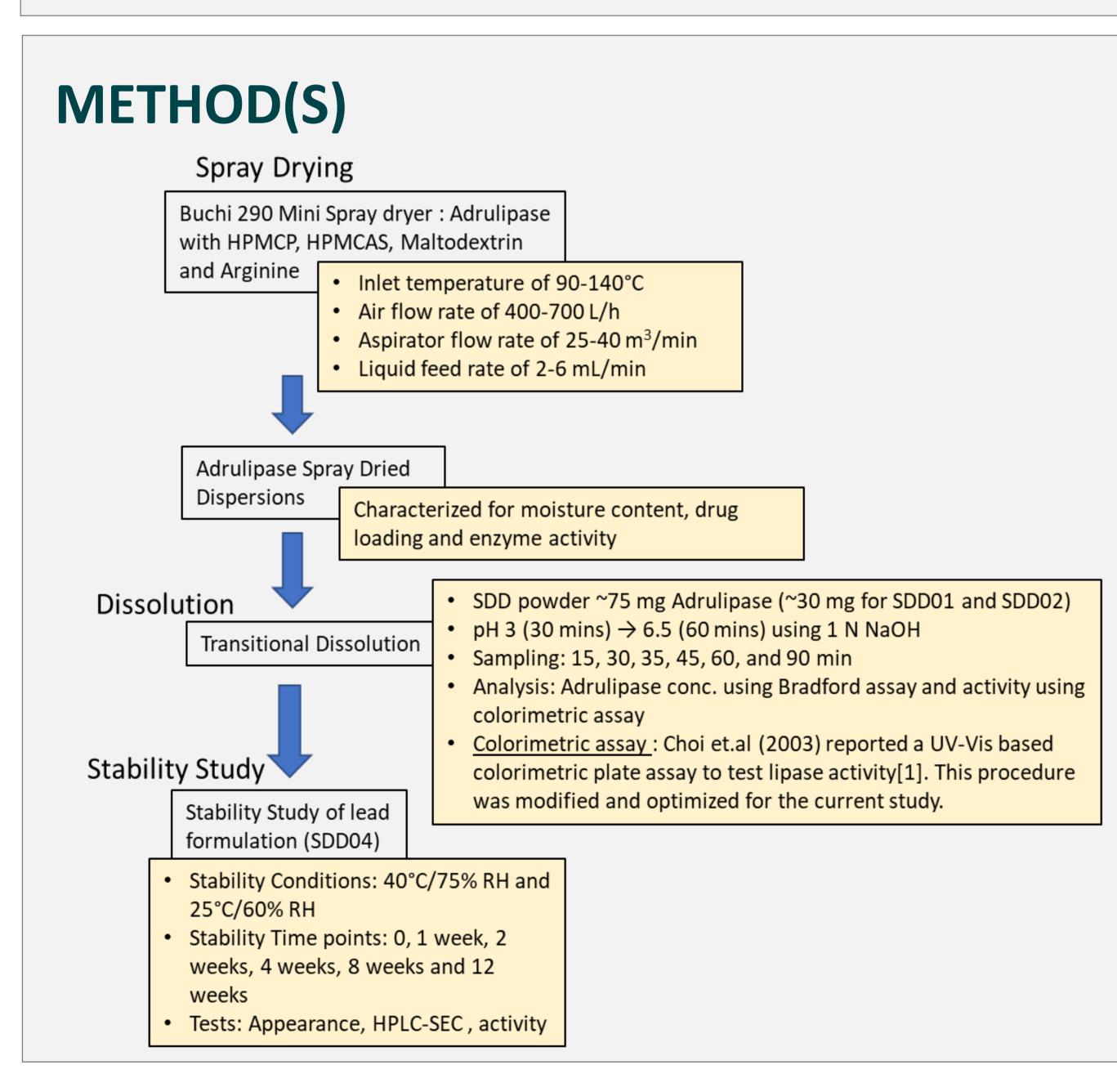
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### PURPOSE

- Oral Delivery of proteins and peptides is challenging due to their po stability in the GI tract (esp. acidic conditions).
- Deficiency of pancreatic lipase leads to a condition called exocrine pancreatic insufficiency (EPI). Administration of porcine pancreatic enzyme replacement therapy (PERT) remains the primary treatment option for patients with EPI.
- Adrulipase, a recombinant non-porcine lipase is currently in development for the treatment of EPI.
- In the present study, several formulations of Adrulipase combined with an enteric polymer, as a spray dried dispersion (SDD), were evaluated to achieve an optimal delayed release profile while retaining Adrulipase activity. A stability study of the selected lead SDD was also conducted.

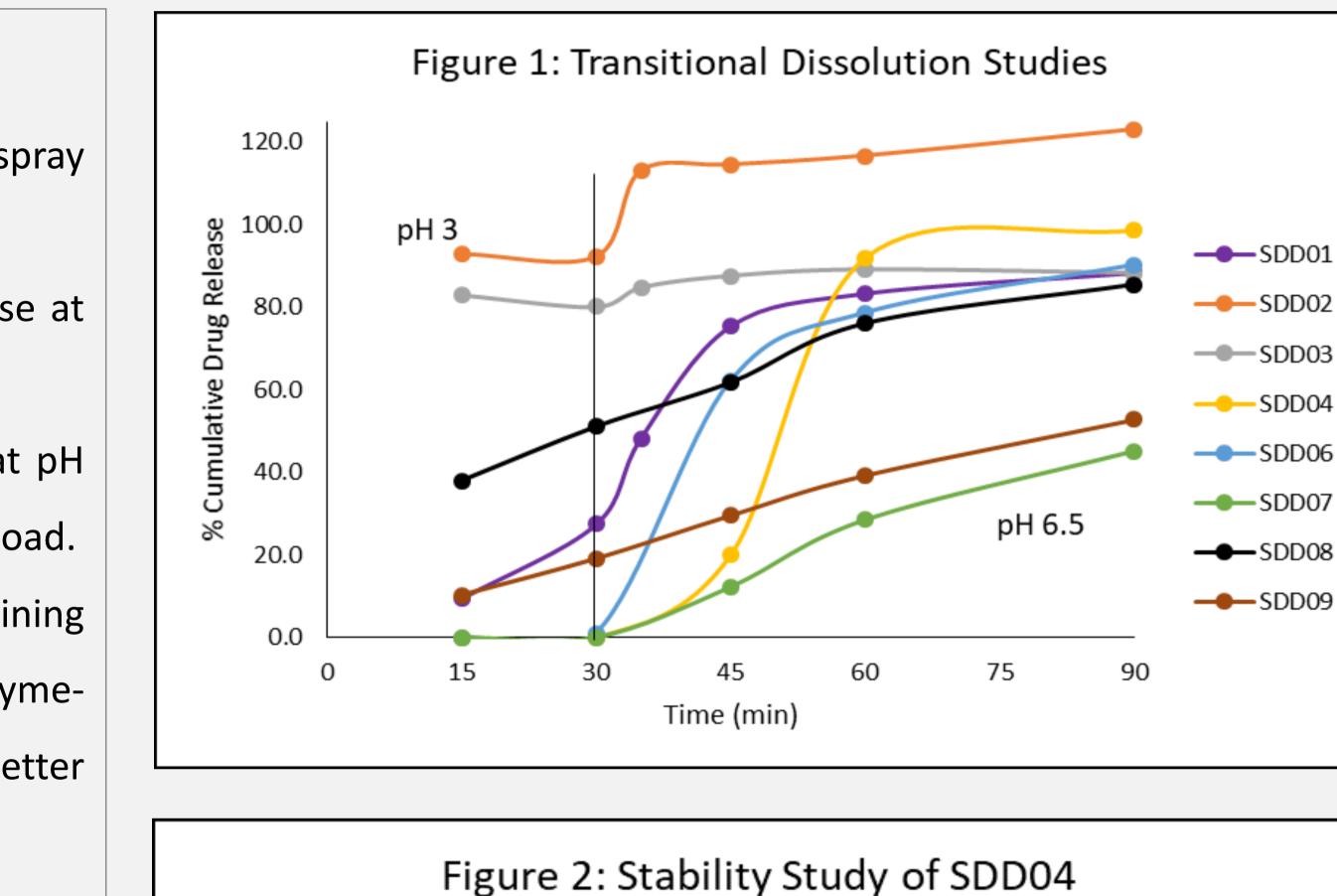
### **OBJECTIVE(S)**

- Evaluate spray dried dispersion formulations of Adrulipase to select a lead formulation optimized for acid protection and dissolution profile for further development.
- To evaluate the stability of lead formulation.



- **RESULT(S)**
- Spray Drying and Transitional Dissolution :
- Adrulipase showed similar enzymatic activity before and after spray drying, with and without an enteric polymer.
- SDD02 and SDD03 contained no enteric polymers (>80% release at pH 3, Fig 1).
- SDD containing arginine (SDD07) showed incomplete release at pH 6.5 versus SDD containing maltodextrin (SDD04) at similar drug load.
- A sticky mass was observed on dissolution of SDDs containing arginine as matrix, which might be due to the formation of enzymearginine agglomerates. Maltodextrin was thus found to be a better stabilizer for Adrulipase compared to arginine.
- API was spray dried with HPMCP only, in an attempt to maximize drug load, however the drug release from the SDD was slow and incomplete (SDD09).
- HPMCP provided better enteric protection than HPMCAS (SDD04) versus SDD08, 0-10% versus ~40% release at pH 3) at similar concentration and drug loading. This might be due to the difference in the structure of drug-polymer-stabilizer matrix formed upon spray drying.
- Lead Formulation :
- SDD04 was selected as the lead for further development of a capsul dosage form.
- The % spray dried yield was approx. 65-70%, contained approximately 50% drug load, was fully protected at pH 3, an retained activity even after 90 minutes under dissolution conditions No significant change in HPLC-SEC peak area as well as activity was observed on storage of the material at both stability condition (Figure 2). No additional impurities/ increase in existing impurities was observed during the testing period.





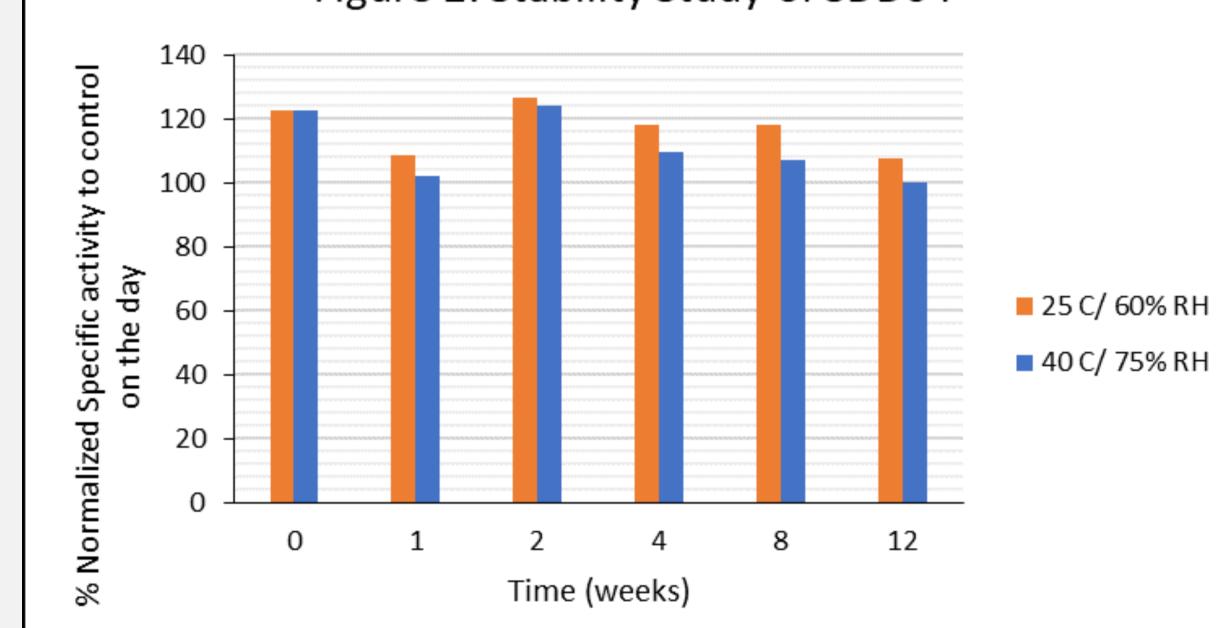


	Table 1 : Composition of SDDs	
le	Name	Composition
	SDD01	API, Maltodextrin, HPMCP
ed	SDD02	API, Maltodextrin
nd	SDD03	API, Maltodextrin
ns.	SDD04	API, Maltodextrin, HPMCP
	SDD05	Spray dried API (no excipients)
vas	SDD06	API ,Arginine, HPMCP
ns	SDD07	API , Arginine, HPMCP
ies	SDD08	API, Maltodextrin, HPMCAS
	SDD09	API, HPMCP

### CONCLUSION(S)

- Nine Adrulipase spray-dried dispersions (SDDs) containing various excipients (HPMCP, HPMCAS, maltodextrin and arginine) were prepared and characterized.
- With the goal to generate drug product with a delayed release profile and the highest possible Adrulipase loading, formulation **SDD04** was selected as the lead formulation for further development.
- **SDD04** showed delayed release profile, offering the best protection at acidic pH and rapid release under intestinal conditions, as well as minor loss of activity over 1) 3-month stability assessment (25°C and 40 °C) and 2) formulation processing activities (granulation).
- Granulation to improve flowability and better retention of enzymatic activity during dissolution has been completed and Investigational Medicinal Product (IMP) is being manufactured for clinical study.
- SDD of enzymes with a stabilizing agent and an enteric polymer may provide a novel approach to enable oral delivery of peptides and proteins as therapeutics.

### REFERENCE

1. Choi, S. J., Hwang, J. M., & Kim, S. I. (2003). A colorimetric microplate assay method for high throughput analysis of lipase activity RNAR Reports 26(A) A17\_A20



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